

# Effect of Long Term $\text{NH}_4^+$ Nutrition on Growth and Yield of Wheat Plants<sup>1</sup>

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## ABSTRACT

The effect of  $\text{NH}_4^+$  nutrition in the development and growth of plants (cv. Klein dorado) was studied in a pot experiment under glasshouse conditions. Plants were grown in 1 kg sand (three plants per pot) and fertilized with a buffered solution containing  $\text{NO}_3^-$  or  $\text{NH}_4^+$  as N-sources. Nitrogen concentration per pot was maintained at  $2.0 \text{ mol} \cdot \text{m}^{-3}$ . Tillering was restrained by manual removal. Plant samples were taken from 30 d after sowing until ripening. Root growth was severely inhibited and shoot growth was markedly depressed by long term  $\text{NH}_4^+$  nutrition. Nevertheless, the specific absorption rate of nitrogen (SARN) was similar in both N treatments until almost two weeks after anthesis. Ear enlargement was accelerated and spikelet differentiated at a faster rate in  $\text{NH}_4^+$  fed plants. However, at ripening, the total number of spikelets per ear, florets per spikelet and grain yield were significantly lower in  $\text{NH}_4^+$  treatment. Total nitrogen concentration in different plant parts did not differ between N treatments, although larger amounts of free  $\text{NH}_4^+$  and total free amino acids were found in the  $\text{NH}_4^+$  treatment during the whole sampling period. Results are discussed in relation to toxic effects due to accumulation of endogenous free ammonium.

Key words: Ammonium toxicity, ear development, free amino acids, *Triticum aestivum* L.

## RESUMEN

Plantas jóvenes de trigo (cv. Klein dorado) fueron cultivadas, en condiciones de invernáculo, en macetas con 1 kg de arena (tres plantas por maceta), las que se fertilizaron periódicamente por medio de un tampón con una solución con  $\text{NO}_3^-$  o  $\text{NH}_4^+$  como fuentes de nitrógeno. Las plantas fueron demacolladas manualmente y la concentración de N por maceta se mantuvo en aproximadamente  $2.0 \text{ mol} \cdot \text{m}^{-3}$ . Se realizaron cosechas periódicas en cada tratamiento desde 30 d después de la siembra hasta la madurez. El tratamiento prolongado con  $\text{NH}_4^+$ , como única fuente de N, redujo el crecimiento de las raíces. No obstante, la tasa específica de absorción de N (SARN) fue similar en ambos tratamientos, hasta casi dos semanas después de la antesis. Tanto el alargamiento del ápice como la diferenciación de las espiguillas se aceleraron en las plantas tratadas con  $\text{NH}_4^+$ , las cuales, en la madurez, presentaron un menor número de espiguillas por espiga, flores por espiguilla y mayor porcentaje de flores fértiles abortadas en comparación con el tratamiento con  $\text{NO}_3^-$ . La concentración de N en las distintas fracciones no difirió significativamente entre ambos tratamientos, aunque las plantas cultivadas con  $\text{NH}_4^+$  acumularon mayores niveles de  $\text{NH}_4^+$  y aminoácidos libres durante el período de muestreo. Los resultados se discuten en relación con los posibles efectos tóxicos provocados por la acumulación de  $\text{NH}_4^+$  libre en los tejidos.

## INTRODUCTION

Different forms of N nutrition distinctly affect both growth and chemical composition of plants (4, 15, 24, 25). Nitrate is the most abundant N sources present in the soil, and the usual N source for the plants. However, ammonium nutrition has often been proposed as an optimum N source for plant growth as it avoids energy wastes due to  $\text{NO}_3^-$  reduction (24). Symptoms of water stress and toxicity have been reported by many authors when  $\text{NH}_4^+$  was used as the sole N source (1, 4, 5, 7). Several factors

have been thought to be responsible for such symptoms, but, unlike the nitrate ion, the accumulation of free  $\text{NH}_4^+$  in plant tissues exerts "per se" toxic effects (13). Nevertheless, many plant species are adapted to  $\text{NH}_4^+$  as the sole N source (24), and some crops like wheat or rice increased their yield when  $\text{NH}_4^+$  was added to a nutrient solution containing  $\text{NO}_3^-$  (9, 28, 29).

Grain production is the main component of crop yield, and all those factors affecting ear development also affect grain number and quality. Several authors have reported increments of grain yield when nitrogen fertilization was increased (10, 11, 18, 27); however there is little information on the effects of different N sources on spike development and grain setting. In this paper, we studied the effects of  $\text{NO}_3^-$  and  $\text{NH}_4^+$  nutrition on growth, ear development and grain yield of wheat plants grown under greenhouse conditions.

<sup>1</sup> Received for publication 16 July 1992.

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## MATERIALS AND METHODS

## Plant culture

Wheat caryopses (*Triticum aestivum* L. cv. Klein Dorado) were germinated on wet tissue paper in the dark. After 72 h, six seedlings were transplanted to plastic pots containing 1 kg of washed sand and periodically irrigated with demineralized water. After a week, plants were reduced to three per pot and fertilization was started. Pots were randomly divided into two groups and the treatments were initiated by fertilizing each pot with 50 ml of a basal solution containing 10.0 mol · m<sup>-3</sup> KNO<sub>3</sub> or 5.0 mol · m<sup>-3</sup> SO<sub>4</sub>(NH<sub>4</sub>)<sub>2</sub> as N sources (Table 1).

Table 1. Nutrient composition of basal fertilization solution.

Macro nutrients	mol m <sup>-3</sup>	Micro nutrients	mmol m <sup>-3</sup>
KCl	4	KCl	50
CaCl <sub>2</sub>	4	H <sub>3</sub> BO <sub>3</sub>	25
KH <sub>2</sub> PO <sub>4</sub>	2	MnSO <sub>4</sub> ·H <sub>2</sub> O	2
FeEDTA 50%	2	ZnSO <sub>4</sub> ·7H <sub>2</sub> O	2
MgSO <sub>4</sub> ·7H <sub>2</sub> O	1	CuSO <sub>4</sub> ·5H <sub>2</sub> O	0.5
		H <sub>2</sub> MoO <sub>4</sub>	0.5

Final nutrient concentration on each pot was 1/5 of the fertilizing solution. Plants were fertilized twice a week during the first 45 days and thrice a week during the following two weeks. Then, both the volume and frequency of fertilization were modified according to plant requirements. Samples of solution percolates from randomly selected pots were periodically analyzed for N content, and their pH recorded during the whole experimental period. When necessary, pots were washed with demineralized water in order to avoid excessive ion accumulation. The pH was periodically adjusted to 6.5 ± 1.0 by addition of H<sub>2</sub>SO<sub>4</sub> or KOH to the treatment solutions.

Tillering was restrained by manual removal so that only the main shoot was allowed to grow. Three pots (i.e., nine plants) per treatment were sampled from 30 d after sowing (d.a.s.) until ripening, at the time intervals shown in the figures. After sampling, roots were carefully washed and rinsed with demineralized water. Plants were divided into roots, leaves plus stems and ears (if present), fresh weights recorded and then samples were freeze dried for further analysis. At ripening, grains were also analyzed as a distinct fraction. Before frying, shoot apices were microdissected and kept in a

mixture of ethanol (96%)-water-formaldehyde-acetic acid (gl.) (5.0:3.5:1.0:0.5) for further observation. The whole experiment was conducted in a glasshouse with daily temperatures of 10 °C - 15 °C at night and 25 °C - 32 °C during the day.

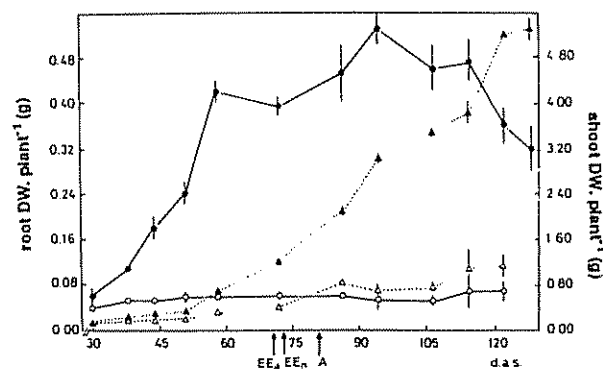
## Plant analysis

Total nitrogen was determined by micro Kjeldahl analysis. Total free amino acids and free ammonium analysis: 0.1 g of freeze dried plant material were extracted with 10 ml of a mixture of ethanol-chloroform-water (12:5:3) overnight at 0 °C. After filtration, 1.5 ml chloroform was added and, after shaking and addition of 2.25 ml of distilled water, the aqueous fraction was extracted. Samples were kept at -18 °C until total partitioning and the aqueous fraction was then isolated for free amino acids and ammonium assays.

Total free amino acids were analyzed through the ninhydrin method (32). Free ammonium was determined by distillation of 0.5 ml aliquots plus 0.5 ml buffer pH 11 (NaOH 1 N: Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub> · 10H<sub>2</sub>O 0.5 M, 1:1). Free NH<sub>4</sub><sup>+</sup> was collected on 2 ml of 2% boric acid and titrated with H<sub>2</sub>SO<sub>4</sub>.

## RESULTS

No differences were observed between plant dry weight in both N treatment at the first sampling. However, NO<sub>3</sub><sup>-</sup> fed plants grew at a faster rate than the NH<sub>4</sub><sup>+</sup>-treated plants so that, at ripening, they had accumulated a 500% higher total dry weight (Fig. 1).

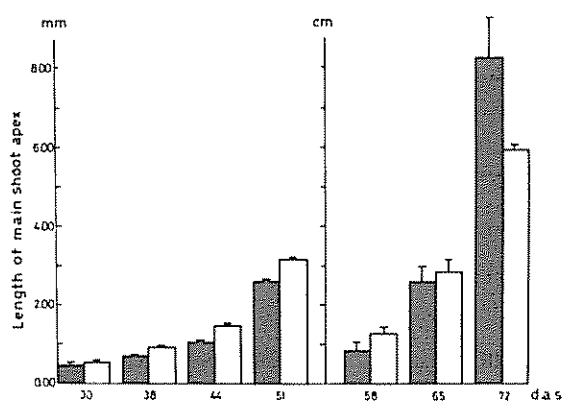


EE = Ear emergence (a= NH<sub>4</sub><sup>+</sup> treatment; n= NO<sub>3</sub><sup>-</sup> treatment). A = Anthesis. Vertical bars represent SE.

Fig. 1. Root dry weight (○) and shoot dry weight (●) of plants supplied with NH<sub>4</sub>-N (open symbols) or NO<sub>3</sub>-N (closed symbols).

Root dry weight in the  $\text{NH}_4^+$  treatment remained almost constant during the whole sampling period, while it largely increased with time until two weeks after anthesis in the  $\text{NO}_3^-$  treated plants (Fig. 1). Shoot growth was also several times faster in the  $\text{NO}_3^-$  fed plants than in the  $\text{NH}_4^+$  treatment.

The ammonium-fed plants showed a faster shoot apex elongation (Fig. 2) accompanied by an earlier spikelet initials differentiation (data not shown). Nevertheless, anthesis occurred at almost the same date in both treatments.



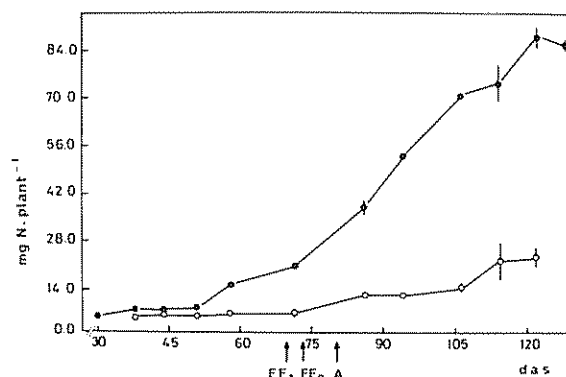
Vertical bars represent SE.

Fig. 2. Length of the main shoot apex from 30 d.a.s. until spike emergence in plants treated with  $\text{NH}_4^+\text{-N}$  (open bars) or  $\text{NO}_3^-\text{-N}$  (closed bars).

Ear maturation occurred at a faster rate under  $\text{NH}_4^+$  nutrition, so that  $\text{NH}_4^+$ -fed plants had completely matured 10 d before  $\text{NO}_3^-$  treatment. At ripening, the number of spikelets per spike and grains per spikelet were significantly lower in  $\text{NH}_4^+$ -fed plants (Table 2). From the average number of florets per spikelet recorded at ear emergence (Table 3), 66.7%, 57.1% and 50.0% of the basal, central and terminal spikelets respectively had aborted in  $\text{NH}_4^+$ -treated plants, while the corresponding values for control treatment were 33.3%, 37.5% and 33.3%. On the other hand, 40% (basal spikelets) to 60% (central and terminal spikelets) of the remaining florets set grain in the  $\text{NO}_3^-$  fed plants, while

no basal florets and only a 33% of the central and terminal spikelets did the same under  $\text{NH}_4^+$  nutrition (Table 2). Thousand-grain weight was also lower in this treatment with respect to that with  $\text{NO}_3^-$  (Table 4).

Total N content per plant was slightly higher in the  $\text{NO}_3^-$  treatment than in the  $\text{NH}_4^+$ -fed plants during the first 20 d of sampling (Fig. 3). However the  $\text{NO}_3^-$  fed plants accumulated nitrogen at a faster rate after this period. As this increment can be related to either a higher plant growth or to a higher N absorption per unit root weight, the specific absorption rate of N (SARN) was estimated in both N treatments (8). SARN did not differ between  $\text{NO}_3^-$  and  $\text{NH}_4^+$ -fed plants until anthesis, although root growth was almost totally inhibited in the latter. The values obtained between 30 and 86 d.a.s. (5 d after anthesis) were 1.64 and 1.69  $\text{mg N} \cdot \text{root DW}^{-1} \cdot \text{d}^{-1}$  respectively. At 94 d.a.s., root growth of  $\text{NO}_3^-$  fed plants began to decline until maturity; however, their SARN almost doubled that of  $\text{NH}_4^+$  treatment during the same period (2.31 vs. 1.19  $\text{mg N} \cdot \text{root DW}^{-1} \cdot \text{d}^{-1}$ ).



EE = Ear emergence (a =  $\text{NH}_4^+$  treatment; n =  $\text{NO}_3^-$  treatment). A = Anthesis Vertical bars represent SE.

Fig. 3. Total nitrogen content in plants supplied with  $\text{NH}_4^+\text{-N}$  (o) or  $\text{NO}_3^-\text{-N}$  (●).

Nitrogen percentage (N%) in shoot and spikes was almost the same under both  $\text{NH}_4^+$  or  $\text{NO}_3^-$  nutrition, and only the roots of  $\text{NH}_4^+$ -treated plants showed a significantly higher N concentration than the  $\text{NO}_3^-$  treatment during the sampling period (Fig. 4).

Table 2. Number of spikelets, florets and grain yield in the mature ear of wheat plants supplied with  $\text{NO}_3\text{-N}$  (data rounded to the nearest digit).

	Florets • Spikelet <sup>1</sup>			Florets which set grain	
	Basal spikelets (2 and 3)*	Central spikelets (8, 9, 10 and 11)	Penultimate and terminal spikelets	Spikelets spike	Grains spike
$\text{NO}_3$	5 SE:0.12	6 SE:0.10	5 SE:0.15	19 SE:0.15	54 SE:0.88
$\text{NH}_4$	2 SE:0.05	3 SE:0.20	3 SE:0.36	16 SE:0.15	8 SE:0.70

SE: Standard error.

(\*): Numbers between brackets stand for those spikelets, in ascending order, which were sampled.

Table 3. Number of florets per spikelet in the ears of wheat plants supplied with  $\text{NO}_3\text{-N}$  or  $\text{NH}_4\text{-N}$  and sampled at ear emergence (data rounded to the nearest digit).

	Basal spikelets (2 and 3)*	Central spikelets (8, 9, 10 and 11)	Penultimate and terminal spikelets
$\text{NO}_3$	8 SE:0.13	9 SE:0.08	8 SE:0.13
$\text{NH}_4$	6 SE:0.13	7 SE:0.11	6 SE:0.27

SE: Standard error

(\*): Numbers between brackets stand for those spikelets, in ascending order, which were sampled.

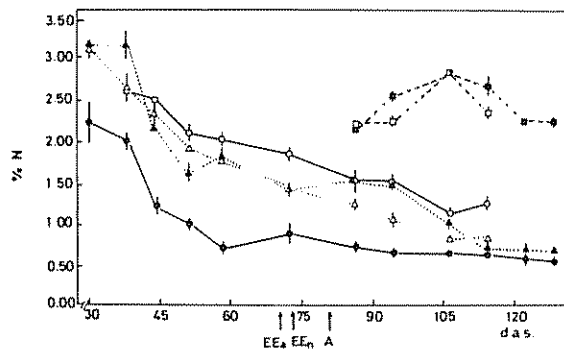
At the first sampling, roots of the  $\text{NH}_4$ -treated plants showed 100% more free ammonium per g DW than roots of the  $\text{NO}_3$ -fed plants (Fig. 5). On the other hand, while ammonium concentration in shoots remained constant in the  $\text{NO}_3$  treatment, it largely increased in the  $\text{NH}_4$ -fed plants, reaching a peak of  $60 \mu\text{mol} \cdot \text{g DW}^{-1}$  at seven days before ripening. At

ripening, free ammonium in shoots had diminished, but was very high in the mature ears of  $\text{NH}_4$ -fed plants in contrast to the levels found in the  $\text{NO}_3$  treatment (Table 4). In both treatments, the concentration of free  $\text{NH}_4$  in the grain fraction accounted for less than 0.5% of that present in the whole spike.

Table 4. Dry weight,  $\text{NH}_4^+$  concentration and total free amino acids concentration in the mature ear and grain fractions of wheat plants supplied with  $\text{NO}_3^-$ -N or  $\text{NH}_4^+$ -N.

Whole ear				
	Dry weight (g)	$\mu\text{mol NH}_4^+$ g DW <sup>-1</sup>	$\mu\text{mol aa}$ g DW <sup>-1</sup>	
$\text{NO}_3^-$	3.76	2.02	31.58	
	SE:0.05	SE:0.78	SE:1.85	
$\text{NH}_4^+$	0.88	82.54	96.37	
	SE:0.12	SE:2.04	SE:2.17	
Grains				
	Dry weight of 1 000 grains (g)	$\mu\text{mol NH}_4^+$ g DW	$\mu\text{mol aa}$ g Dw	N % grain
$\text{NO}_3^-$	30.30	0.00 Sc: 0.00	0.00 SE: 0.00	3.24 SE: 0.04
$\text{NH}_4^+$	20.84	0.70 SE: 0.70	8.87 SE: 2.88	3.1 SE: 0.08

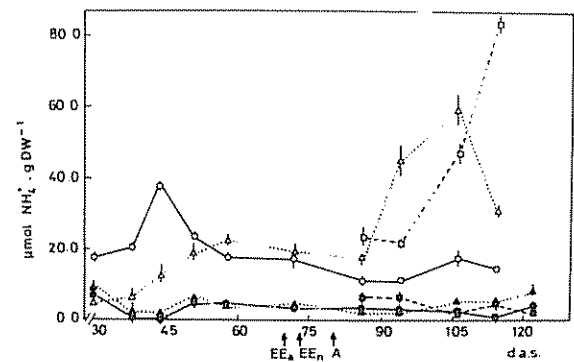
SE: standard error



EE = Ear emergence (a=  $\text{NH}_4^+$  treatment; n=  $\text{NO}_3^-$  treatment).  
A = Anthesis. Vertical bars represent SE.

Fig. 4. Nitrogen percent in root (o), shoot ( $\Delta$ ) and ear ( $\square$ ) of plants supplied with  $\text{NH}_4^+$ -N (open symbols) or  $\text{NO}_3^-$ -N (closed symbols).

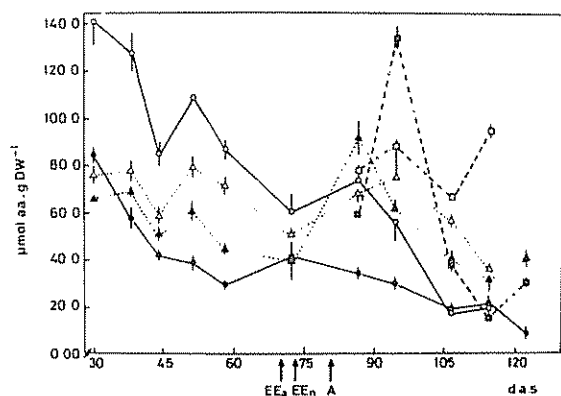
The concentration of free amino acids in roots was initially 60% higher in  $\text{NH}_4^+$ -treated plants, and in both N treatments it decreased with time (Fig. 6). The concentration of free amino acids in shoots was initially lower than that present in the roots, and only slight differences were found between N treatments until ear emergence. In the spike, it increased during the first



EE = Ear emergence (a=  $\text{NH}_4^+$  treatment; n=  $\text{NO}_3^-$  treatment).  
A = Anthesis. Vertical bars represent SE.

Fig. 5. Free ammonium concentration in root (o), shoot ( $\Delta$ ) and ear ( $\square$ ) of plants supplied with  $\text{NH}_4^+$ -N (open symbols) or  $\text{NO}_3^-$ -N (closed symbols).

week after ear emergence, especially in those plants fed with  $\text{NO}_3^-$ -N. However, while in this case it rapidly decreased after that time, it remained constant in  $\text{NH}_4^+$  treatment. From the content of total free amino acids present in mature ears, only 6% ( $\text{NH}_4^+$  treatment) and 0% ( $\text{NO}_3^-$  treatment) were in the grain fraction (Table 4).



EE = Ear emergence (a = NH<sub>4</sub>-N treatment; n = NO<sub>3</sub>-N treatment).  
A = Anthesis. Vertical bars represent SEE.

Fig. 6. Total free amino acids concentration in root (o), shoot (△) and ear (□) of plants supplied with NH<sub>4</sub>-N (open symbols) or NO<sub>3</sub>-N (closed symbols).

## DISCUSSION

The causes of the toxic effects of the ammonium ion on higher plants have been widely reviewed (13, 24), but not aspects are completely clear. Impaired plant growth, high shoot: root ratios and premature senescence have been reported for young wheat plants (9, 12) and other species (1, 6, 31) when NH<sub>4</sub>-N was supplied at increasing external concentrations or during a prolonged period. Mehrer and Mohr (22) have recently described these symptoms as the "ammonium toxicity syndrome".

While in several cases the observed effects could be explained as a consequence of different metabolic changes due to NH<sub>4</sub>-N uptake and assimilation, it is known that the endogenous accumulation of free ammonium is "per se" deleterious to plant growth (5, 13, 17, 30).

In the present experiment, plant growth was severely inhibited by ammonium nutrition in both shoots and roots, (Fig. 1) and later the ear (Tables 2 and 3). However, the organ most severely affected was the root, since it slightly grew only slightly during the whole sampling period under NH<sub>4</sub>-N nutrition (Fig. 1), with a consequent increase in the shoot:root ratio. This impairment of root growth may be a consequence of the high free ammonium concentration (Fig. 5). However, the absorption capacity of the root did not seem to have

been affected, as the SARN from 30 d a s. until almost two weeks after anthesis were similar in both N treatments.

An estimation of the N translocation during 30 d to 86 d after sowing showed that the ammonium-fed plants had transported almost 100% more mg N per day and per mg of root DW than the NO<sub>3</sub>-N fed plants (data not shown). This would indicate that shoot growth under NH<sub>4</sub>-N nutrition was not initially impaired because of a deficient N translocation. The high levels of free NH<sub>4</sub>-N present in both root and shoot tissues of plants supplied with NH<sub>4</sub>-N suggest that toxic effects due to an excessive accumulation of free NH<sub>4</sub>-N, rather than N deficiency, were responsible for the observed symptoms.

Most of the NH<sub>4</sub>-N absorbed by the roots is assimilated within the roots via the glutamine synthetase-glutamate synthase pathway, so that glutamine and other amino acids are the N derivatives transported to the shoot (2, 3, 19). As the root plays a key role in the process of NH<sub>4</sub>-N detoxification, the high levels of NH<sub>4</sub>-N found under our experimental conditions suggest that the capacity for ammonium assimilation of the root system has been largely overcome (Fig. 5). The fact that NH<sub>4</sub>-N assimilation was occurring is suggested by the high concentration of free amino acids present both in root and shoot tissues (Fig. 6), although NH<sub>4</sub>-N accumulation could also have affected protein synthesis (17).

Reduced shoot growth, accelerated apex development and leaf senescence are symptoms observed under water stress or nitrogen stress (8, 21, 23, 27), so that they are not specific responses to NH<sub>4</sub>-N, but general responses of plants to stress. However, root growth was severely diminished under NH<sub>4</sub>-N nutrition rather than stimulated, as it is often observed under water or N stress (14, 16).

On the other hand, while N% in the grain fraction did not differ between treatments (Table 4), both grain setting and grain filling were markedly diminished in those plants with high endogenous concentrations of free NH<sub>4</sub>-N (Tables 2 and 4).

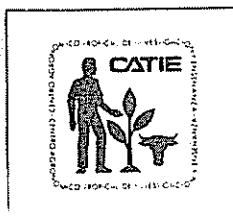
Apex development and ear emergence were initially accelerated by ammonium feeding (Fig. 2). Similar results have been previously reported for wheat plants (20), though the physiological underlying mechanism is still unknown.

It can be concluded that, under  $\text{NH}_4^+$  nutrition, while the plant becomes older and the shoot:root ratio changes as a consequence of a highly impaired root growth, the  $\text{NH}_4^+$  detoxification capacity of the plant decreases, thus increasing the ion concentration in the leaves and ears to toxic levels.

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## CENTRO AGRONÓMICO TROPICAL DE INVESTIGACION Y ENSEÑANZA (CATIE)

Anuncia la apertura del período de admisión para su Programa de Maestría en Ciencias Agropecuarias y Recursos Naturales, año académico 1995-1996:

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| <p><b>I. Maestría en Sistemas de Producción Agrícola Sostenible, con énfasis en:</b></p> <ul style="list-style-type: none"> <li>a. Cultivos Tropicales</li> <li>b. Fitoprotección</li> <li>c. Sistemas Agroforestales</li> </ul> | <p><b>II. Maestría en Manejo Integrado de los Recursos Naturales, con énfasis en:</b></p> <ul style="list-style-type: none"> <li>a. Manejo de Cuencas Hidrográficas</li> <li>b. Manejo y Conservación de la Biodiversidad</li> <li>c. Manejo y Silvicultura de Bosques Tropicales</li> </ul> |
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### Requisitos:

- Poseer título universitario en áreas afines.
- Superar proceso de admisión (examen y evaluación curricular).
- Menor de 35 años de edad (de preferencia).
- La admisión implica el ser considerado estudiante-asistente de investigación, durante su permanencia en el CATIE.

### Fechas importantes para el año 1994

30 de abril:	Límite para la recepción de solicitudes
7 de junio:	Límite para la recepción de notas universitarias y documentos personales
3ra. semana de junio:	Examen de admisión (simultáneo en todos los países)
31 de agosto:	Comunicación de resultados del proceso de admisión
10 de noviembre:	Comunicación de asignación de becas administradas por el CATIE
30 de noviembre:	Límite de aceptación de beca por parte de los estudiantes
9 de enero de 1995:	Inicio de los cursos

Para mayor información, diríjase a:

CATIE  
Programa de Enseñanza para el Desarrollo y la Conservación  
Turrialba, Costa Rica - Apartado 7170  
Teléfonos: (506) 556-1016, 556-6431 / Fax: (506) 556-1533